Food Science and Technology Research, **27** (3), 359–367, 2021 copyright©2021, Japanese Society for Food Science and Technology doi: 10.3136/fstr.27.359

Original Paper



Distribution and diffusion coefficients of glucose and sodium chloride onto konjac and egg white gel

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Received November 17, 2020 ; Accepted February 23, 2021

The sorption isotherms and diffusion coefficients of glucose and sodium chloride on konjac and egg white gel were measured at temperatures ranging from 4 °C to 80 °C to systematically organize the transport of seasoning compounds during food manufacturing. The distribution coefficients, defined as the slopes of the sorption isotherms, decreased as the temperature increased. The enthalpy changes calculated from the temperature dependence of the distribution coefficients for glucose and sodium chloride on konjac and egg white gel were approximately -2 kJ/mol. The effective diffusion coefficients ranged from 10^{-10} to 10^{-8} m²/s and increased with the temperature. The activation energies of diffusion in konjac and egg white gel were approximately 17 and 9 kJ/mol, respectively; glucose and sodium chloride had similar activation energies for diffusion. Diffusion in multi-component systems was also studied; the degrees of sorption in multi-component systems.

Keywords: distribution coefficient, effective diffusion coefficient, egg white gel, glucose, konjac, sodium chloride

Introduction

The penetration of seasoning compounds, such as saccharides and salts, into foodstuffs can alter the texture and taste; therefore, great attention is being paid to the seasoning transport phenomena during food manufacturing. However, although many processes that depend on experience and intuition are performed, few are conducted efficiently. The development of a method for the prediction and control of seasoning transport phenomena from a scientific viewpoint will contribute to both large-scale food manufacturing and small-scale domestic cooking.

To understand the transport phenomena of seasonings in foods, analyses of the distribution equilibrium and diffusion rates are required. Although prior studies have indicated the amounts of typical seasoning compounds that are sorbed onto potatoes (Take and Otsuka, 1968) and whale meat (Take and Otsuka, 1973), and the amounts of sodium chloride that are sorbed onto starch gel, egg white gel, and eight types of vegetables (Matsuzaki *et al.*, 1971), these studies did not assess the sorption equilibrium, and only evaluated the interim penetration progress during cooking. Other studies have determined the diffusion rates of sodium chloride in radish (Hashiba *et al.*, 2007), potato (Hashiba *et al.*, 2013), cheese (Morris *et al.*, 1985; Turhan and Kaletunç, 1992), pork (Wood, 1966; Graiver *et al.*, 2006), and vegetables (Kusnadi and Sastry, 2012). Since the experimental temperatures varied from low (Wood, 1966; Turhan and Kaletunç, 1992; Graiver *et al.*, 2006) to high (Hashiba *et al.*, 2007; Hashiba *et al.*, 2013), it is difficult to compare the results of these studies. Moreover, because of the complexity of ingredient compositions and food microstructures, it is difficult to apply the findings of one study to an arbitrary manufacturing process.

In this study, the systematic organization of seasoning transport phenomena in foodstuffs was evaluated from multiple viewpoints. Firstly, simple gel-like materials, namely, egg white gel and konjac, were used as model foods with and without an electrical charge, respectively. Secondly, sodium chloride and glucose were adopted as models of lowmolecular-weight seasonings with and without an electrical charge, respectively. The main purpose of this research was to

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本論文は Food Science and Technology Research, 27(3), 359-367 (2021) 掲載論文を転載したものである.

obtain systematic knowledge about the sorption equilibrium and diffusion rates of seasonings in foods using model compounds with different electrostatic properties.

Materials and Methods

Materials Commercially available konjac (Co-op Kobe, Kobe, Japan) and soy sauce (Kikkoman, Chiba, Japan) were purchased from a local supermarket in Kobe. Dried egg white powder (type K, No. 10) was obtained from Kewpie Egg (Tokyo, Japan). D(+)-glucose, sodium chloride, liquid paraffin, and a glucose CII-test kit were purchased from FUJIFILM Wako Pure Chemical Corporation (Osaka, Japan).

Preparation of egg white gel Dried egg white powder (type K, No. 10) was dissolved in seven times its weight of ion-exchanged water. The solution was centrifuged using a centrifugal separator (himac CR 21GII; Koki Holdings, Tokyo, Japan) at 12 000 rpm (approximately 18 800 \times g) for 15 min at 4 °C to obtain the supernatant. The absorbance of the supernatant diluted 500 times and measured at 280 nm using a U-2000 spectrophotometer (Hitachi, Tokyo, Japan) or a Cary 3500 UV-Vis spectrophotometer (Agilent Technologies Japan, Tokyo, Japan) was approximately 0.23, and this value did not change remarkably in repeated experiments.

The obtained supernatant was poured into a plastic cup (LS115-200T, 114.5 mm in width × 114.5 mm in depth × 25.7 mm in height; Tokan Kogyo, Tokyo, Japan) to a depth of approximately 15 mm, and heated in a steam bath for 20 min. A T-fal steam cooker (Groupe SEB Japan, Tokyo, Japan) or a hand-made steamer was used for heating. When the egg white solution was directly poured into the plastic cup, the obtained heat-set gel adhered to the plastic cup, and it was difficult to separate the gel from the cup without causing a collapse. To avoid this problem, a light coating of liquid paraffin was applied to the cup before the pouring of the egg white solution to improve the mold-releasability of the egg white gel after heating. The procedure of coating paraffin onto the mold when preparing the egg white gel was adopted from a previous report (Nakagawa et al., 1994). After the egg white gel was separated from the cup, the five sides of the gel that were in contact with the walls and bottom of the cup were trimmed by 5 mm; the trimmings were not used for any of the experiments. The obtained gel is hereafter referred to as the egg white gel.

Moisture content of konjac and egg white gel The moisture contents of konjac and egg white gel were measured by drying them at 135 °C for 5 h using a DV600 constanttemperature oven (Yamato Scientific Co., Ltd., Tokyo, Japan). The weight change after drying was calculated and regarded as the moisture content.

Measurement of the distribution coefficient Model food samples (konjac and egg white gel) were cut into cubes of approximately 10 mm. Five grams of these samples were precisely weighed and soaked in glucose and sodium chloride solutions, the concentrations of which ranged from 10 to 100 kg/m³ and from 3 to 150 kg/m³, respectively. The volume of the solution was approximately 80 mL, and it was replaced several times at appropriate time intervals. After soaking for more than 20 h, the konjac or egg white gel was removed from the solution. The glucose and sodium chloride that had sorbed onto the model foods were extracted and analyzed using the methods described below. To measure the distribution coefficients, a CSH-110 thermostat chamber (Espec, Osaka, Japan) was used to maintain the experimental temperature at 25 °C, 40 °C, 50 °C, or 60 °C, and a URD-50MRF refrigerator (Fukushima Galilei, Osaka, Japan) and an NTB-221 programmable precision bath (Tokyo Rikakikai, Tokyo, Japan) were used to maintain the temperature at 4 °C and 80 °C, respectively.

Determination of the effective diffusion coefficient The effective diffusion coefficients of glucose and sodium chloride in konjac and egg white gel were determined by estimating the temporal increase in the sorbed amounts. Briefly, 5 g of sample cubes were soaked in a solution containing 10 kg/m³ of glucose or sodium chloride, as described above. The sorbed amounts were estimated after an appropriate time interval using the procedure mentioned in the previous section. To determine the maximum sorption amounts of glucose and sodium chloride, we analyzed the sorbed amounts after 6 h.

To measure the effective diffusion coefficients, the experimental temperature was maintained at 25 °C and 60 °C using the CSH-110 thermostat chamber along with an MLR-350H plant growth chamber (PHC Holdings, Tokyo, Japan) at 5 °C and a Viscomate VM-150F bath (Toki Sangyo, Tokyo) at 80 °C.

Measurement of the distribution coefficient in multicomponent systems Three different methods were adopted to investigate the sequence of the sorption of seasoning compounds. Model foods were soaked in either a glucose and sodium chloride solution, glucose and sodium chloride solution after equilibration in glucose solution, or glucose and sodium chloride solution after equilibration in sodium chloride solution. In this experiment, the concentrations of both glucose and sodium chloride were fixed at 10 kg/m³. To achieve sorption equilibrium, the model foods were soaked in the respective solutions for more than 20 h during each soaking process. The other conditions were the same as those described in the previous sections.

Extraction of glucose and sodium chloride from the model foods The model foods (konjac or egg white gel) were ground using a mortar and pestle, and 50 mL of ion-exchanged water was added to the mortar. After 30 min, the mixture was filtered using No. 2 filter paper (Toyo Roshi Kaisha, Tokyo, Japan). The obtained filtrate was diluted to a volume of 50 mL,

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and used for the analysis of the glucose and sodium chloride contents.

Analysis of glucose and sodium chloride Glucose and sodium chloride concentrations were estimated using the glucose CII-test kit and a SAT-500 salt analyzer (DKK-Toa, Tokyo, Japan), respectively. For the glucose assay, the absorbance of the reaction mixture was measured at 505 nm using the U-2000 spectrophotometer or the Cary 3500 UV-Vis spectrophotometer. When using the SAT-500 salt analyzer, the injection volume of each sample was 200 µL.

Results and Discussion

Heating conditions during the preparation of the egg white gel The temperature of the center of the egg white solution during heating was above 75 °C for the last 5 min, and it was heated until it finally reached 85 °C. The atmospheric temperature in the steam bath was above 95 °C during heating. It has been reported that coagulation and precipitation of the proteins occur in egg white when the egg white solution is heated at 74 °C for 5 min (Akkouche et al., 2012). We supposed that heating in a steam bath above 95 °C for 20 min was sufficient to complete the gelation of the egg white solution.

It is well-known that the heating conditions of egg white solution, such as the temperature and duration, affect the obtained heat-set egg white gel (Doi and Kitabatake, 1989). In this experiment, instead of studying the effects of heating conditions on the properties of egg white gel, we employed a method of egg white gel preparation that repeatedly yielded gels with constant properties. Thereafter, we analyzed the experimental data assuming that all of the experiments were performed using egg white gels with the same properties.

Moisture content of the konjac and egg white gel The moisture contents of the konjac and egg white gel, measured using the drying method, were 96.1% \pm 0.1% and 89.6% \pm 0.2% (w. b.), respectively (n = 5). The solid contents of the konjac and egg white gel were 3.9% and 10.4% (w. b.), respectively. Although the solid content of the egg white gel was twice that of konjac, the concentration used was deemed necessary because of the poor gelation properties of egg white at lower concentrations.

Temperature dependence of the distribution coefficients In preliminary experiments, we investigated the temporal changes in the amounts of glucose sorbed onto konjac at 25 °C for 1, 2, 4, 6, and 20 h (data not shown). We found that the amount of sorbed glucose did not change between 6 and 20 h. To ensure equilibrium, we employed a soaking time of 20 h to measure the distribution coefficients.

The sorption isotherms of glucose and sodium chloride onto konjac and egg white gel were measured over a temperature range of 4 °C to 80 °C, and some representative

60 40 20 0 20 40 60 80 100 0 Concentration [kg/m³] Fig. 1. Sorption isotherms of glucose onto konjac

(empty symbols) and egg white gel (filled symbols) at 4 °C (circle), 25 °C (triangle), 60 °C (square), 80 °C (diamond); results are expressed as average \pm standard deviation (n = 3).

isotherms are shown in Fig. 1. The selection criterion for egg white gel and konjac as model foods was the presence and absence of an electrical charge on their skeleton molecules, respectively, as mentioned in the Introduction section. All of the sorption isotherms, including those shown in Fig. 1, were linear. Assuming that Henry's law could be applied to the sorption of glucose and sodium chloride onto konjac and egg white gel, Eq. 1 was employed to define the distribution coefficient of the seasoning compounds onto the model foods, $K_D [m^3/kg]$.

$$v = K_{\rm D} \times C$$
 Eq. 1

where $C [kg/m^3]$ is the concentration in the liquid phase, and q [kg/kg] is the amount of sorbed seasoning compound. The initial concentrations of both glucose and sodium chloride in konjac and egg white gel were 0 kg/kg.

q

Fig. 2 shows the temperature dependence of the distribution coefficient. As shown in the figure, the distribution coefficients decreased with increasing temperature. The distribution coefficients of glucose and sodium chloride on konjac were similar at a given temperature, but the distribution coefficients of sodium chloride on egg white gel were lower than those of glucose. This difference in the distribution coefficients on egg white gel at certain temperatures may be attributable to the electrostatic repulsion between egg white protein and sodium chloride, which does not occur between egg white gel and glucose.

The enthalpy change of the distribution, ΔH [J/mol], and the entropy change, ΔS [J/(mol·K)], were calculated from the





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Table 1. Enthalpy change and entropy change of sorption					
	Konjac		Egg white gel		
	Glucose	Sodium chloride	Glucose	Sodium chloride	
Enthalpy change [kJ/mol]	-2.4	-2.5	-2.2	-1.6	
Entropy change [J/(mol·K)]	-10.5	-10.6	-9.3	-9.6	

Temperature [°C] 60 50 40 30 20 10 100 80 0 e^0 Distribution coefficient $\times 10^3$ [m³/kg] $e^{-0.1}$ é -0.3 -0.4 $e^{-0.5}$ $e^{-0.6}$ $e^{-0.7}$ $e^{-0.8}$ 0.9 2.8 3.0 3.2 3.4 3.8 2.6 3.6 10³/T [1/K]

Fig. 2. Temperature dependence of glucose (circle) and sodium chloride (triangle) distribution coefficients onto konjac (empty symbols) and egg white gel (filled symbols); results are expressed as average \pm standard deviation (n = 5-8).

temperature dependence of the distribution coefficients by applying the Van 't Hoff equation (Eq. 2).

where R [J/(mol·K)] is the gas constant, and T [K] is the absolute temperature.

The calculated values of ΔH and ΔS are listed in Table 1. The values of ΔH were independent of the combination of the model food and seasoning compound, and ranged from -2.5 to -1.6 kJ/mol. The values of ΔS ranged from -10.6 to -9.3 J/(mol·K). The changes in Gibbs free energy calculated at each experimental temperature from the values of ΔH and ΔS in Table 1 were all positive irrespective of the experimental temperature. These positive changes in Gibbs free energy were inconsistent with the sorption occurring spontaneously; this might have resulted from the fact that the ΔH values obtained in our study were relatively low when compared to that representing an ion exchange interaction between crystalline antimonic (V) acid and alkali metal ions, such as sodium and potassium ions, which has been reported to be about -25 kJ/mol (Abe, 2001). The relatively weak temperature dependence of the distribution coefficients might have caused the low values of the estimated ΔH , which could not offset the negative values of ΔS . The low values of ΔH also suggest that when glucose and sodium chloride permeate into konjac and egg white gel, they do not interact strongly with the gel skeleton, but simply penetrate the porous regions in the gel.

Temperature dependence of the effective diffusion coefficients The temporal changes in the amounts of glucose and sodium chloride sorbed onto konjac and egg white gel were measured to estimate the effective diffusion coefficients. The temporal changes in the amounts of glucose sorbed onto konjac and egg white gel at 25 °C and 60 °C are shown in Fig. 3. By applying Eq. 3 and the calculation methods described in previous reports (Crank, 1975; Nakanishi *et al.*, 1977; Odake *et al.*, 1990; Sakai, 1991) to the experimental data, the effective diffusion coefficient, $D_{\rm eff}$ [m²/s], was calculated for each experiment. The solid curves in Fig. 3 show the lines calculated using Eq. 3 and the estimated $D_{\rm eff}$ values, which were in good agreement with the experimental values.

$$\frac{q_{\infty}-q_{t}}{q_{\infty}-q_{0}} = 8\sum_{m=0}^{\infty}\sum_{n=0}^{\infty}\sum_{p=0}^{\infty}\frac{\exp\left[-\{(m+1/2)^{2}+(n+1/2)^{2}+(p+1/2)^{2}\}\pi^{2}D_{\text{eff}}t/l^{2}\right]}{(m+1/2)^{2}(n+1/2)^{2}(p+1/2)^{2}\pi^{6}}$$
..... Eq. 3

where l [m] (5 × 10⁻³ m) is the characteristic length of konjac or egg white gel, t [s] is time, q_0 and q_t [kg/kg] are the sorbed amounts at time 0 and t, respectively, and q_{∞} [kg/kg] is the maximum sorbed amount. Letters m, n, and p express all integers larger than 0 used in the sigma calculation.

The temperature dependence of the calculated $D_{\rm eff}$ values is shown in Fig. 4. The $D_{\rm eff}$ values ranged from 10^{-10} to 10^{-8} m²/s, with the same order of magnitude as those reported based on previous studies on other foods (Morris *et al.*, 1985; Turhan and Kaletunç, 1992). In the same model food, the $D_{\rm eff}$ values of sodium chloride tended to be higher than those of glucose, which reflects its smaller molecular size and faster diffusion in food, as shown by the Stokes–Einstein equation (Edward, 1970; Young *et al.*, 1980).

As shown in Fig. 4, the D_{eff} values increased with the temperature. By applying the Arrhenius equation (Eq. 4) to the

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10-1

 10^{-2}

0

0.2

 $(q_{\infty}-q_{t})/(q_{\infty}-q_{0})$

 $(t/l^2) \times 10^9 \,[s/m^2]$ Fig. 3. Temporal changes in glucose sorption onto konjac (empty symbols) and egg white gel (filled symbols) at 25 °C (circle) and 60 °C (triangle); solid lines represent lines calculated using Eq. 3 and estimated $D_{\rm eff}$ values.

0.4

С

0.6

С

0.8

1.0



Fig. 4. Temperature dependence of effective diffusion coefficients of glucose (circle) and sodium chloride (triangle) in konjac (empty symbols) and egg white gel (filled symbols).

temperature dependence of D_{eff} , the activation energy, E_{a} [J/mol], was calculated.

Table 2. Activation energy of diffusion

	Activation energy [kJ/mol]		
	Glucose	Sodium chloride	
Konjac	16.2	17.5	
Egg white gel	8.1	9.8	

where A [m²/s] is the frequency factor. The calculated E_a values are presented in Table 2. The E_a values of glucose and sodium chloride did not differ greatly in the same model food, but those of both glucose and sodium chloride in konjac were larger than those in egg white gel. In other words, diffusion in konjac is more sensitive to temperature changes than diffusion in egg white gel. The diffusion temperature dependence of seasoning compounds in model foods may be influenced by the type of food rather than by the type of seasoning compound.

A comparison of Fig. 2 and Fig. 4 showed that the equilibrium sorbed amounts of seasoning compounds became lower at higher temperatures, whereas the diffusion rate was higher at higher temperatures. We employed distribution coefficients as representative values of the sorption equilibrium, and analyzed them using the Van 't Hoff equation (Eq. 2). The lower values of the distribution coefficients at higher temperatures resulted in negative values of enthalpy change, as shown in Table 1. In contrast, the temperature dependence of the diffusion coefficients was considered to obey the Arrhenius equation (Eq. 4). The diffusion coefficient increases with the temperature, because the value of activation energy in Eq. 4 is always positive. The results shown in Fig. 4 are consistent with this rule.

The $E_{\rm a}$ values for several seasoning compounds have been reported. For example, the E_a values of sodium chloride in potatoes are approximately 20 to 24 kJ/mol (Liu, 1992), whereas those in white cheese are approximately 23 to 30 kJ/mol (Turhan and Kaletunç, 1992). Although the Ea values obtained in this study for konjac and egg white gel were lower than previously reported values, this could be attributed to the structural and compositional complexity of the sample foods, such as the presence of cell walls in potatoes or emulsioncontaining milk fat in cheese. Changes in the value of D_{eff} at high temperatures are considered to be due to both an increase in the molecular diffusion coefficient of glucose or sodium chloride and a change in the pore size of the model food. Compared to potatoes (Liu, 1992) and white cheese (Turhan and Kaletunç, 1992), the microstructure of konjac and egg white gel may not be complex. Hence, the temporal changes in the microstructure of the model foods in our research might not have been as drastic as those of complex foods, although some intermolecular bonds might have formed between egg white proteins, as will be discussed below. This could be the reason why the activation energy of diffusion was smaller than

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that in previous reports, because the change in the value of $D_{\rm eff}$ was mainly due to a change in the molecular diffusion coefficient.

Relationship between the effective diffusion and molecular diffusion coefficients Fig. 5 shows the relationship between the molecular diffusion coefficients in the dilute solution $(D_0 [m^2/s])$ and the effective diffusion coefficients (D_{eff} [m²/s]) of glucose and sodium chloride in each model food. The values of the molecular diffusion coefficients of glucose were calculated using Eq. 5, which was derived from reported data (Ribeiro et al., 2006) by applying the Arrhenius equation. The molecular diffusion coefficients of sodium chloride at each temperature were calculated from the temperature dependence represented by Eq. 6, as reported elsewhere (Caldwell, 1973).

$$D_{0, \text{ Glucose}} = 1967.4 \times \{ \exp[-2.379 \times (10^{3} / T)] \} / 10^{9}$$

$$\cdots \qquad \text{Eq. 5}$$

$$D_{0, \text{ NaCl}} = \{ 0.44 + 0.0423 \times (T - 273.15) \} / 10^{9}$$

$$\cdots \qquad \text{Eq. 6}$$

where $D_{0, \text{ Glucose}}$ and $D_{0, \text{ NaCl}}$ [m²/s] are the molecular diffusion coefficients in dilute solutions of glucose and sodium chloride, respectively.

In Fig. 5, four points with the same symbols represent the results at 5 °C, 25 °C, 60 °C, and 80 °C from the left of the horizontal axis. As shown in Fig. 5, the effective diffusion coefficients (D_{eff}) of glucose in konjac and egg white gel (empty and filled circles, respectively) and sodium chloride in egg white gel (filled triangles) are lower than the molecular diffusion coefficients (D_{0}). Furthermore, the D_{eff} values of sodium chloride in konjac (empty triangles) are slightly higher than the D_0 values. The apparent inconsistency between the estimated D_{eff} of sodium chloride in konjac and the cited D_0 may be because the precise measurement of D_0 is much more difficult than that of D_{eff} .

Although there is a good correlation between the experimental D_{eff} values of glucose and sodium chloride in konjac and the cited D_0 values ($R^2 > 0.95$), the D_{eff} and D_0 for egg white gel exhibit a weak correlation ($R^2 < 0.70$). The slope of the straight line in Fig. 5 is called the hindering factor, which accounts for the structural effect of the gel and chemical interactions between the gel and solutes (Lamberg, 1990; Varzakas and Tzia, 2014). The results shown in Fig. 5 suggest that the hindering factor of egg white gel is affected by temperature changes. In other words, the structure of egg white gel and solutes may change with the temperature. Shimizu *et al.* (1991) reported that when transparent ovalbumin gel was heated below 100 °C and cooled repeatedly, intermolecular



Fig. 5. Relationship between effective diffusion coefficients and molecular diffusion coefficients of glucose (circle) and sodium chloride (triangle) in konjac (empty symbols) and egg white gel (filled symbols); four different points with same symbols represent results obtained at four different temperatures (5, 25, 60, and 80 °C).

disulfide bridges formed, resulting in the polymerization of the denatured ovalbumin. Although the preparation method of egg white gel in this work was different, the observed change in the hindering factor might have been because of the formation of similar intermolecular covalent bonds at high temperatures.

Distribution coefficients in multi-component systems To demonstrate the practical applicability of the model system to manufacturing processes, the distribution coefficients in multi-component systems were measured. Fig. 6 shows the relationship of the distribution coefficients in single- and binary-component systems containing glucose and/or sodium chloride.

In this study, three different methods were adopted to investigate the sequence of sorption of seasoning compounds, namely, model foods were soaked in either glucose and sodium chloride solution (shown as circles in Fig. 6), glucose and sodium chloride solution after equilibration in glucose solution (shown as triangles in Fig. 6), or glucose and sodium chloride solution after equilibration in sodium chloride solution (shown as rectangles in Fig. 6). In Fig. 6, the symbols represent the distribution coefficients of sodium chloride on egg white gel (0.61), sodium chloride on konjac (0.73), glucose on konjac (0.74), and glucose on egg white gel (0.82) from the left of the horizontal axis.

The three kinds of symbols (circles, triangles, and rectangles) show similar values for each combination of model food and seasoning compound, which suggests that the Distribution of glucose and NaCl onto konjac and egg white gel



Fig. 6. Relationship between distribution coefficients in single- and binary-component systems; symbols represent distribution coefficients of sodium chloride onto egg white gel (0.61), sodium chloride onto konjac (0.73), glucose onto konjac (0.74), and glucose onto egg white gel (0.82), from the left of the horizontal axis, when soaked in glucose and sodium chloride solution (circle), glucose and sodium chloride solution after equilibration in glucose solution (triangle), and glucose and sodium chloride solution after sodium chloride solution equilibration in (rectangle), respectively.

sequence of glucose and sodium chloride sorption does not affect the total sorbed amounts in the binary-component system. Moreover, for the sorption of glucose onto konjac, sodium chloride onto konjac, and sodium chloride onto egg white gel, the distribution coefficients were similar in the single- and binary-component systems. In contrast, for the sorption of glucose onto egg white gel, the distribution coefficients in the binary-component system were approximately 0.2 units lower than those in the singlecomponent system. In other words, the amount of glucose sorbed onto egg white gel decreased when sodium chloride coexisted in the liquid phase. Moreover, the decrease in the amount of glucose sorbed onto egg white gel in the binary system was not affected by the sequence of sorption of glucose and sodium chloride, as the three soaking patterns showed similar values.

Only the values of the distribution coefficients of glucose on egg white gel in the binary system decreased. Nakamura *et al.* (1958) reported that the amount of saccharides adsorbed onto an anion-exchange resin decreased when sodium chloride was added to the saccharide solution. They concluded that the decrease in the adsorbed amount was because of ion exchange between the counter anion in the resin and the chloride anion in solution. Taking their report into account, the decrease in the distribution coefficient of glucose on egg white gel in the binary system was due to a similar ion exchange between the ions in egg white gel and sodium or chloride ions.

We also compared single- and multi-component systems using soy sauce as the liquid phase. The amounts of sodium chloride sorbed onto konjac and egg white gel were measured at 25 °C. Sodium chloride solutions with different concentrations and commercially available soy sauce were used as the liquid phase. The concentration of sodium chloride in soy sauce was determined to be 174 kg/m³. When sodium chloride solution was used as the liquid phase, the sodium chloride distribution coefficients (average ± standard deviation) on konjac and egg white gel were $0.73 \times 10^{-3} \pm 0.03$ \times 10^{-3} m^3/kg and 0.61 \times 10^{-3} \pm 0.03 \times 10^{-3} m^3/kg, respectively (Fig. 2). In contrast, when soy sauce was used as the liquid phase, the sodium chloride distribution coefficients in konjac and egg white gel were $0.52 \times 10^{-3} \pm 0.00 \times 10^{-3}$ m³/kg (n = 3) and $0.41 \times 10^{-3} \pm 0.01 \times 10^{-3} \text{ m}^3/\text{kg}$ (*n* = 3), respectively, which were lower than when sodium chloride solution was used. Nanbu et al. (1995a, 1995b, 1997) reported that the amounts of sorbitol sorbed onto walleye pollock in the presence and absence of sodium chloride varied considerably, whereas those of sodium chloride were not affected by the presence of sorbitol. They attributed the changes in the amounts of sorbed sorbitol to fish denaturation proteins and the polymerization of myosin heavy chains during sodium chloride penetration. Other researchers have also reported that when soy sauce is used as the liquid phase, temporal changes in the amount of sodium chloride and monosodium glutamate sorbed onto potatoes and whale meat are higher than those when a single component solution of sodium chloride or monosodium glutamate is used (Take and Otsuka, 1968; Take and Otsuka, 1973). They concluded that the increase in the amounts of sorbed sodium chloride and monosodium glutamate might have been because of the synergistic effect of these two seasoning compounds. In our research, opposite results were obtained. It is known that synthetic resins are used in the decoloring process of soy sauce (Furuta, 1967). The residues of konjac and egg white gel after extraction with ionexchanged water were slightly brown in color, indicating that the color pigment originating from soy sauce had moved into the konjac and egg white gel. Odake (2010) reported that soy sauce pigment moved into hanpen (Japanese fish cake) after 24 h of soaking. As mentioned previously, sodium or chloride ions may be exchanged with the ions in egg white gel. The amounts of sorbed sodium chloride can decrease when the color pigment in soy sauce is competitively adsorbed onto the

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ionic section of egg white gel. Both our study and previous reports suggest that when several kinds of seasoning compounds penetrate foodstuffs simultaneously, interactions between the seasoning compounds and food as well as between the compounds themselves play important roles.

Conclusions

The sorption isotherms of glucose and sodium chloride on konjac and egg white gel were linear at temperatures ranging from 4 °C to 80 °C. The enthalpy changes calculated from the temperature dependence of the distribution coefficients, which are defined as the slopes of the sorption isotherms, remained nearly constant regardless of the combination of the model food and seasoning compound. The estimated effective diffusion coefficients increased with the temperature. The activation energies of glucose and sodium chloride diffusion were similar in each model food. When both glucose and sodium chloride coexisted in the liquid phase, their distribution coefficients differed from when only glucose or sodium chloride existed in the liquid phase.

Nomenclature

- A =frequency factor [m²/s]
- C =concentration in liquid phase [kg/m³]
- D_0 = molecular diffusion coefficient in dilute solution [m²/s]
- D_{0, Glucose} = molecular diffusion coefficient in dilute solution of glucose [m²/s]
- D_{0, NaCl} = molecular diffusion coefficient in dilute solution of sodium chloride [m²/s]

 $D_{\rm eff}$ = effective diffusion coefficient [m²/s]

 $E_a = activation energy [J/mol]$

 $\Delta H = \text{enthalpy change [J/mol]}$

 $K_{\rm D}$ = distribution coefficient [m³/kg]

l = characteristic length [m]

m, n, p = integers larger than 0 used in the sigma calculation (Eq. 3) [-]

q =sorbed amount [kg/kg]

- $q_0 =$ sorbed amount at time 0 [kg/kg]
- q_t = sorbed amount at time t [kg/kg]
- q_{∞} = maximum sorbed amount [kg/kg]
- $R = \text{gas constant} [J/(\text{mol} \cdot K)]$
- $\Delta S = \text{entropy change } [J/(\text{mol}\cdot K)]$

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t = time [s]
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T = absolute temperature [K]

References

- Abe, M. (2001). Fundamental experiments of ion-exchange reaction for the researchers. *J. Ion Exch.* (*Nippon Ion Kokan Gakkaishi*). **12**, 20–29. (in Japanese)
- Akkouche, Z., Aissat, L. and Madani, K. (2012). Effect of Heat on egg white proteins. In "International Conference on

Applied Life Sciences," ed. by F. Nejadkoorki. IntechOpen, Turkey, pp. 407–413.

- Caldwell, D. R. (1973). Thermal and Fickian diffusion of sodium chloride in a solution of oceanic concentration. *Deep-Sea Res. Oceanogr. Abstr.*, **20**, 1029–1039.
- Crank, J. (1975). "The Mathematics of Diffusion." 2nd ed." Clarendon Press, Oxford, U.K.
- Edward, J. T. (1970). Molecular volumes and the Stokes-Einstein equation. J. Chem. Educ., 47, 261–270.
- Doi, E., and Kitabatake, N. (1989). Structure of glycinin and ovalbumin gels. *Food Hydrocol.*, 3, 327–337.
- Furuta, T. (1967). Decoloring of soy sauce. Journal of the brewing society of Japan (Nippon Jouzoukyoukai Zasshi).
 62, 1309–1313.
- Graiver, N., Pinotti, A., Califano, A., and Zaritzky, N. (2006).
 Diffusion of sodium chloride in pork tissue. *J. Food Eng.*, 77, 910–918.
- Hashiba, H., Komiyama, J., Nakanishi, T., and Gocho, H. (2007). Dual mode diffusion of NaCl in Japanese radish under cooking conditions. *J. Food Sci.*, **72**, C154–C162.
- Hashiba, H., Gocho, H., and Komiyama, J. (2013). Dual mode diffusion and sorption of sodium chloride in pre-cooked potato (*Solanum tuberosum* L.). J. Food Res., 2, 66–79.
- Kusnadi, C., and Sastry, S. K. (2012). Effect of temperature on salt diffusion into vegetable tissue. *Int. J. Food Prop.*, **15**, 1148–1160.
- Lamberg, I. (1990). Transport phenomena in potato tissue. A study with applications to blanching, drying and frying. Ph.D. Thesis, University of Lund, Lund, Sweden.
- Liu, H. (1992). A kinetic study of salt diffusion in potato at high temperature. *Int. J. Food Sci. Technol.*, 27, 443–455.
- Matsuzaki, A., Ehara, A., Yamazaki, Y., and Matsumoto, F. (1971). Experimental study on the permeation of salt in the food. J. Home Econ. Jpn. (.Kaseigaku Zasshi). 22, 227–231. (in Japanese)
- Morris, H. A., Guinee, T. P., and Fox, P. F. (1985). Salt diffusion in Cheddar cheese. J. Dairy Sci., 68, 1851–1858.
- Nakagawa, Y., Ohta, T., Nakatsuka, M., Tanaka, T., Hayashi, Y., Matsumura, Y., and Mori, T. (1994). Physical properties of EGG white gels modified by addition of powdered chitin. J. Jpn. Soc. Food Sci. Technol. (Nippon Shokuhin Kogyo Gakkaishi). 41, 321–326. (in Japanese)
- Nakamura, M., Yoshihiro, Y., and Higurashi, K. (1958). Sorption of saccharides on ion-exchange resin. *The Journal* of the Society of Chemical Industry, Japan (Kogyo Kagaku Zasshi). 61, 1150–1152. (in Japanese)
- Nakanishi, K., Adachi, S., Yamamoto, S., Matsuno, R., Tanaka, A., and Kamikubo, T. (1977). Diffusion of saccharides and amino acids in cross-linked polymers. *Agric. Biol. Chem.*, 41, 2455–2462.
- Nanbu, S., Kiuchi, H., Yamamoto, Y., Kawamori, Y., Funatsu,

Distribution of glucose and NaCl onto konjac and egg white gel

- Y., and Arai, K. (1995a). Effect of sorbitol on change in myofibrillar protein of meat block from walleye pollack during soaking in NaCl solution. *Bulletin of the Japanese Society of Fisheries (Nippon Suisan Gakkaishi)*. **61**, 225–231. (in Japanese)
- Nanbu, S., Kiuchi, H., Yamamoto, Y., Kawamori, Y., Funatsu, Y., and Arai, K. (1995b). Effect of sorbitol on change in myofibrillar protein of cured meat block from walleye pollack during dehydration. *Nippon Suisan Gakkaishi*, **61**, 233–241. (in Japanese)
- Nanbu, S., Kiuchi, H., Ohishi, A., Kitajima, T., and Arai, K. (1997). Effects of NaCl and sorbitol permeations into meat from walleye pollack on denaturation of myofibrillar proteins and moisture content during soaking. *Bulletin of the Japanese Society of Fisheries (Nippon Suisan Gakkaishi)* 63, 608–615. (in Japanese)
- Odake, S., Hatae, K., Shimada, A., and Iibuchi, S. (1990). Apparent diffusion coefficient of sodium chloride in cubical agar gel. *Agric. Biol. Chem.*, **54**, 2811–2817.
- Odake, S. (2010). Science of cooking and salt. Presented at 8th Saltscience Symposium, Tokyo, September 28, pp. 17–22. (in Japanese)
- Ribeiro, A. C. F., Ortona, O., Simões, S. M. N., Santos, C. I.
 A. V., Prazeres, P. M. R. A., Valente, A. J. M., Lobo, V. M.
 M., and Burrows, H. D. (2006). Binary mutual diffusion coefficients of aqueous solutions of sucrose, lactose, glucose, and fructose in the temperature range from (298.15 to 328.15) K. J. Chem. Eng. Data, 51, 1836–1840.
- Sakai, M. (1991). Transfer rate of glucose in fish flesh. J. Jpn. Soc. Food Sci. Technol. (Nippon Shokuhin Kogyo Gakkaishi). 38, 303–308. (in Japanese)

- Shimizu, A., Kitabatake, N., Higasa, T., and Doi, E. (1991). Melting of the ovalbumin gels by heating: reversibility between gel and sol. J. Jpn. Soc. Food Sci. Technol. (Nippon Shokuhin Kogyo Gakkaishi). 38, 1050–1056.
- Take, T., and Otsuka, H. (1968). Studies on the scientific cookings of various foodstuffs (Part 1) on the diffusibility of various seasonings into cooked potatoes. *Kaseigaku Zasshi*, **19**, 326–328. (in Japanese)
- Take, T., and Otsuka, H. (1973). Scientific cookings of various foods (Part 2) diffusibility of various seasonings into cooked whale meat. *Chouri Kagaku*, 6, 125–129. (in Japanese)
- Turhan, M., and Kaletunç, G. (1992). Modeling of salt diffusion in white cheese during long-term brining. J. Food Sci., 57, 1082–1085.
- Varzakas, T., and Tzia, C. (2014). "Food Engineering Handbook: Food Engineering Fundamentals." CRC Press, FL, U.S., p. 270.
- Wood, F. W. (1966). The diffusion of salt in pork muscle and fat tissue. J. Sci. Food Agric., 17, 138–140.
- Young, M. E., Carroad, P. A., and Bell, R. L. (1980). Estimation of diffusion coefficients of proteins. *Biotechnol. Bioeng.*, 22, 947–955.